

BIOLOGICAL PROPERTIES OF  
STREPTONIGRIN DERIVATIVES

I. ANTIMICROBIAL AND  
CYTOCIDAL ACTIVITIES

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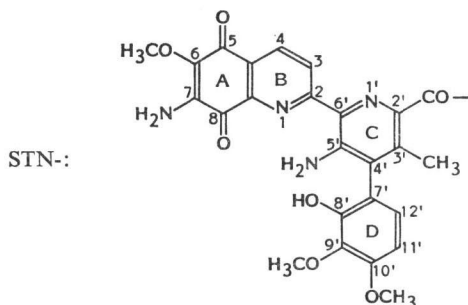
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In the course of our screening for inhibitors  
against reverse transcriptase of avian myelo-  
blastosis virus, two inhibitors have been isolated  
from the cultured broth of *Streptomyces re-*

Table 1. Structures of streptonigrin derivatives.



1	STN-OH	
2	STN-OCH <sub>3</sub>	
3	STN-NH <sub>2</sub>	A
4	STN-NHCH <sub>2</sub> CH <sub>2</sub> OH	A, B
5	STN-NHCH <sub>2</sub> CH <sub>2</sub> CH <sub>2</sub> N(CH <sub>3</sub> ) <sub>2</sub>	B
6	STN-NHCH <sub>2</sub> COOH	B*
7	STN-NHCH <sub>2</sub> COOCH <sub>3</sub>	B
8	STN-NHOH	B
9	STN-NHNH <sub>2</sub>	B
10	STN-NHNHCONH <sub>2</sub>	A, B
11	STN-NHNHCSNH <sub>2</sub>	A, B
12	STN-NH(CH <sub>2</sub> ) <sub>3</sub> NH(CH <sub>2</sub> ) <sub>4</sub> NH-STN	B
13	STN-NH(CH <sub>2</sub> ) <sub>3</sub> NH(CH <sub>2</sub> ) <sub>4</sub> NH(CH <sub>2</sub> ) <sub>3</sub> NH-STN	B

\* 6 was obtained by hydrolysis of 7.

Table 2. Antimicrobial spectra of streptonigrin derivatives.

Test microbe	MIC (μg/ml)						
	1	2	3	4	5	8	11
<i>Staphylococcus aureus</i> FDA 209 P	0.78	>25	>25	>25	>25	25	>25
<i>S. aureus</i> Smith	0.39	>25	25	>25	>25	3.12	6.25
<i>Micrococcus flavus</i> FDA 16	6.25	>25	>25	>25	>25	>25	>25
<i>M. luteus</i> PCI 1001	1.56	>25	>25	>25	>25	>25	>25
<i>Bacillus anthracis</i>	<0.05	>25	6.25	12.5	12.5	1.56	6.25
<i>B. subtilis</i> PCI 219	<0.05	25	>25	>25	25	3.12	6.25
<i>Corynebacterium bovis</i> 1810	0.78	>25	25	>25	>25	>25	>25
<i>Escherichia coli</i> K-12	3.12	>25	>25	>25	>25	>25	>25
<i>E. coli</i> K-12 ML 1629	6.25	>25	>25	>25	>25	>25	>25
<i>E. coli</i> NIHJ	0.39	>25	>25	>25	>25	>25	>25
<i>Shigella sonnei</i> 191-66	3.12	>25	>25	>25	>25	>25	>25
<i>Proteus vulgaris</i> OX 19	6.25	>25	>25	>25	>25	>25	>25
<i>Klebsiella pneumoniae</i> PCI 602	<0.05	25	>25	12.5	25	3.12	3.12
<i>Salmonella typhosa</i> T-63	6.25	>25	>25	>25	>25	>25	>25
<i>Serratia marcescens</i>	0.78	>25	>25	>25	>25	>25	>25
<i>Candida tropicalis</i> NI 7495	25	>25	>25	>25	>25	>25	>25
<i>C. pseudotropicalis</i> NI 7494	25	>25	>25	>25	>25	>25	>25
<i>C. albicans</i> 3147	>25	>25	>25	>25	>25	>25	>25
<i>Saccharomyces cerevisiae</i>	>25	>25	>25	>25	>25	>25	>25

Antimicrobial spectra of 8 and 9 were the same. 10 and 13 showed the same antimicrobial spectra with that of 4. 5 and 12 had the same antimicrobial spectrum. 6 and 7 did not inhibit the growth of above microbes at a concentration of 25 μg/ml.

*trostaticus*<sup>11</sup>. One of them was identified with streptonigrin, the aminoquinolinequinoid antibiotic produced by *S. flocculus*<sup>2-4</sup>, and the other was proved to be novel and named re-trostatin<sup>11</sup>. Streptonigrin (**1**) showed strong antitumor activity but the clinical application of **1** was limited by its extraordinary strong side effects mainly due to bone marrow depression<sup>5,6</sup>. Chemical modifications of the amino group on C7 or C5', or the hydroxyl group on C8' of **1** usually resulted in substantial loss of the antitumor activity<sup>7</sup>. For example, the *in vitro* and

*in vivo* antitumor activities of isopropylidene azastreptonigrin were reported to be approximately 0.01% and 1.0%, respectively, of those of **1**<sup>8</sup>. While streptonigrin methyl ester (**2**) showed marked antitumor activity by parental administration as well as **1**, the maximum tolerated dose of **2** for humans was 5~6 times that of **1**<sup>9,10</sup>. Therefore, the carboxyl group on C2' of **1** was modified by the acid chloride method using acid chloride of **1** as an intermediate (Method A) or by the reaction of an amine with **1** in the presence of phenyl bis(2-

Fig. 1. Effects of streptonigrin derivatives on the growth of L5178Y/S and L5178Y/ADR cells.

A test sample dissolved in DMSO (2.5 mg/ml) was diluted with serum-free FISCHER's medium to make a test solution. A mixture of the test solution (0.2 ml) and the cell suspension ( $5.0\sim 6.0 \times 10^4$  cells/1.8 ml) in FISCHER's medium containing 10% horse serum (Gland Island Biological Co.) was incubated in a tightly capped test tube at 37°C for 72 hours<sup>12</sup>.

The cell numbers were counted by using a hemocytometer after the incubation.  
A, L5178Y/S; B, L5178Y/ADR.

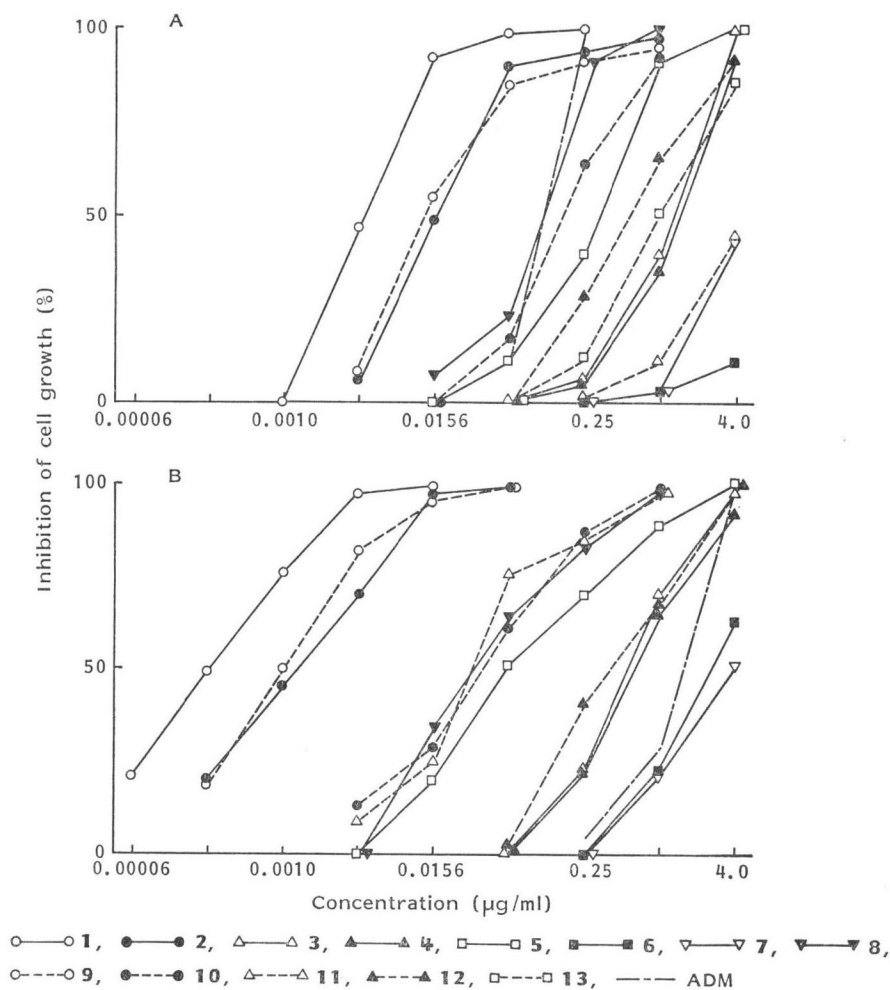


Fig. 2. Effects of streptonigrin derivatives on the growth of P388/S and P388/ADR cells.

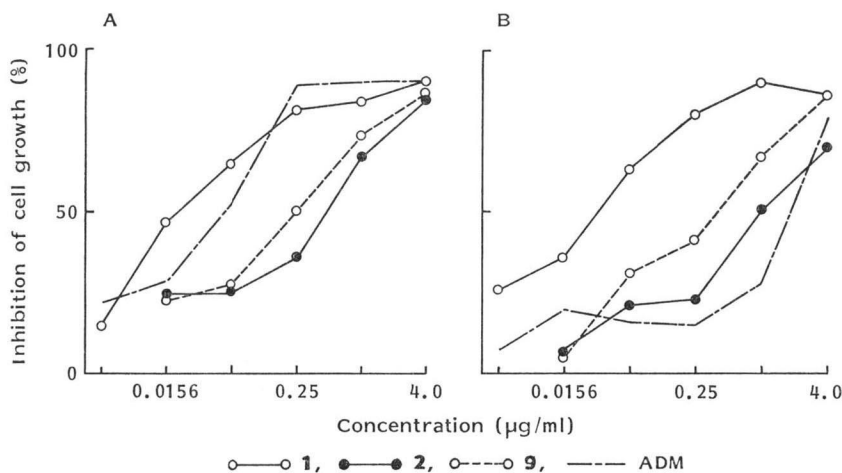
Roswell Park Memorial Institute Medium 1640 supplemented with 10% fetal calf serum (GIBCO), 10  $\mu\text{M}$  2-hydroxyethyl disulfide (Aldrich Chemical Co.) and kanamycin (100  $\mu\text{g}/\text{ml}$ ) was used as a culture medium.

P388/S and P388/ADR ascites cells were harvested from the peritoneal cavity of each tumor-bearing DBA/2NCrj mouse 6 days after transplantation and subcultured twice at 37°C in Falcon No. 1007 plastic dishes (Falcon Plastics) in a humidified atmosphere of 5% CO<sub>2</sub>.

A mixture of the test solution (0.1 ml) prepared as described in the legend to Fig. 1 and the cell suspension (5.0  $\times 10^4$  cells/0.9 ml) in the medium was cultured in a loosely capped test tube at 37°C for 48 hours<sup>14</sup>.

The cell numbers were counted in a Model Dn Coulter counter.

A, P388/S; B, P388/ADR.



thioxo-1,3-thiazolidine-3-yl)phosphine oxide (Method B)<sup>11</sup>. Biological properties of **1** and its derivatives (**2**~**13**, Table 1) are described in this paper.

Antimicrobial spectra of the streptonigrin derivatives were determined by the agar dilution method on glucose nutrient agar. Streptonigrin hydroxamic acid (**8**) and streptonigrin hydrazide (**9**) showed rather weak antimicrobial activities when compared with that of **1** as seen in Table 2. The other derivatives showed very weak or faint activities as exemplified by lack of activity observed for **2**, **6** or **7** at a concentration of 25  $\mu\text{g}/\text{ml}$ .

Growth inhibitory activities of the streptonigrin derivatives against various tumor cells were determined using a parental line of lymphosarcoma L5178Y cells (L5178Y/S), an adriamycin (ADM)-resistant subline of L5178Y cells (L5178Y/ADR)<sup>12,13</sup>, a parental line of P388 leukemia cells (P388/S) and an ADM-resistant subline of P388 cells (P388/ADR)<sup>14-17</sup>. **1**, **2** and **9** showed 32, 8 and 8 times, respectively, stronger cytotoxicities than ADM against

L5178Y/S (Fig. 1). Furthermore, it is worthwhile to emphasize that the ID<sub>50</sub> of **1**, **2** and **9** against L5178Y/S (4.0  $\times 10^{-3}$ , 1.6  $\times 10^{-2}$  and 1.3  $\times 10^{-2}$   $\mu\text{g}/\text{ml}$ , respectively, Fig. 1A) are remarkably higher than those against L5178Y/ADR (2.7  $\times 10^{-4}$ , 1.3  $\times 10^{-3}$  and 0.9  $\times 10^{-3}$   $\mu\text{g}/\text{ml}$ , respectively, Fig. 1B). In fact, NISHIMURA *et al.*<sup>12</sup> observed that none of the tested antitumor agents showed higher cytotoxicity against L5178Y/ADR than against L5178Y/S. The sensitivity of L5178Y/ADR to **1** and its derivatives is apparently higher than that of L5178Y/S. The collateral sensitivity of this type has been reported mainly in *in vivo* tumor systems<sup>16,15</sup>. INABA *et al.*<sup>10</sup> observed, however, the *in vitro* collateral sensitivity of 6-mercaptopurine-resistant sublines of P388 and L1210 leukemia to the purine antagonists, 5-carbamoyl-1H-imidazol-4-yl piperonylate and 4-carbamoylimidazolium-5-olate. The ID<sub>50</sub> of **1**, **2**, **9** and ADM against P388/S were 1.6  $\times 10^{-2}$ , 5.0  $\times 10^{-1}$ , 2.5  $\times 10^{-1}$  and 6.0  $\times 10^{-2}$   $\mu\text{g}/\text{ml}$ , respectively (Fig. 2A). Though P388/ADR had been reported to show wide cross-resistance against various antitumor

agents of different groups<sup>16,17</sup>, no significant difference was observed between the ID<sub>50</sub> of **1**, **2** and **9** against P388/S and the corresponding values of P388/ADR as shown in Fig. 2. According to the previous findings by INABA *et al.*<sup>14,15</sup>, P388/ADR acquired resistance to a variety of antitumor agents due to the enhanced active efflux of these substances, and **1**, **2** and **9** seemed not to be affected to the common efflux system of P388 leukemia cells.

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